

SOP No: AHT 30 (a)

SUBJECT: Retro-Orbital Sinus Bleed in the Mouse

POLICY: Mice must be bled under anaesthetic
Operator must be experienced and competent in this
technique.
Bleeding may only be performed once unless otherwise
approved by the AEC.

PRECAUTIONS: Gloves, eye protection, long sleeve gown, closed in shoes.

EQUIPMENT: anaesthetic
Capillary tubes
Eppendorf tubes

PROCEDURE:

1. Anaesthetise mouse (anaesthetic of choice-specify dose rate, route of injection and volume.)
2. Restrain the animal so that the skin around the eye pulls tight and the eye bulges.
3. Gently push the exposed end of the capillary tube behind the eyeball into the eye socket at the outer edge of the eye.
4. Tilt the mouse so that it is facing the floor, belly down. Allow blood to flow down the tube.
5. Expect to collect no more than 200µl/bleed. If multiple bleeds have been approved, do not bleed again for at least 14 days.
6. Remove excess blood from around the eye.

RECOMMENDATIONS:

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REFERENCES

1. Perspectives on Animal Use Biological Effects of Blood Loss: Implications for Sampling Volumes and Techniques. In: ILAR News (1989), 31(4).
2. Removal of blood from laboratory mammals and birds: First report of the BVA/FRAME/RSPCA/UFAW Joint working group on refinement. Lab Anim (1993) 27, 1-22.
3. J. Donovan and P. Brown. Blood Collection. In: Current Protocols in Immunology, (eds: A. Kruisbeek et al), John Wiley & Sons, New York, NY, USA, Unit 1.7.