

- SOP No:** AHT 01
- SUBJECT:** Handling and restraint of laboratory mice (*Mus spp*)
- POLICY:** Completion of UQ small animal handling workshop or equivalent and adequate training provided by qualified animal facility staff.
- PRECAUTIONS:** Gown/lab coat, gloves, facemask, closed in shoes.

Laboratory Animal Allergens (LAA)

It is important that all precautions are taken to minimize exposure to laboratory animal allergens. Use Personnel Protective Equipment (PPE) provided and inform a supervisor or animal facility manager of known allergies to laboratory animals.

If bitten, do not make sudden movements or jerk the hand away. Place the mouse down gently in a secure cage, the mouse should let go and move away. Ensure the cage lid is secure, remove gloves and place them in the appropriate waste bin. Wash the hands thoroughly with soap and water and dry with paper towel.

Hygiene

Before and after handling mice, always wash hands, to remove odours from other species or blood which can be distressing to mice. Hands can also spread infections from one animal to another. If bitten, wash the affected area immediately and apply antiseptic. Complete and submit the required OH&S pro forma.

- EQUIPMENT:** Appropriate surfaces for handling mice (e.g. cage wire), appropriate rodent cages.

- PROCEDURE:**
1. **Hand scoop** (scoop mouse up with one or both hands).



- 2. Tail hold (Lift mouse up by the base of the tail using thumb and forefinger).**



- 3. Tail hold with support (as with tail hold but with one hand supporting the mouse's body).**



- 4. Transfer mouse in plastic tube or enrichment device. When transferring mice from one cage to another, wait for the mouse to run into the plastic tube/enrichment device and use this to transfer the animal.**

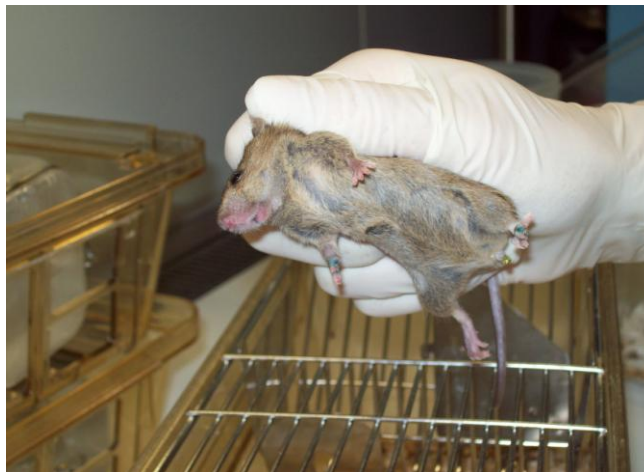


5. Manual restraint:

Pick up mouse by base of the tail and place onto easy to grip surface such as a wire cage lid.

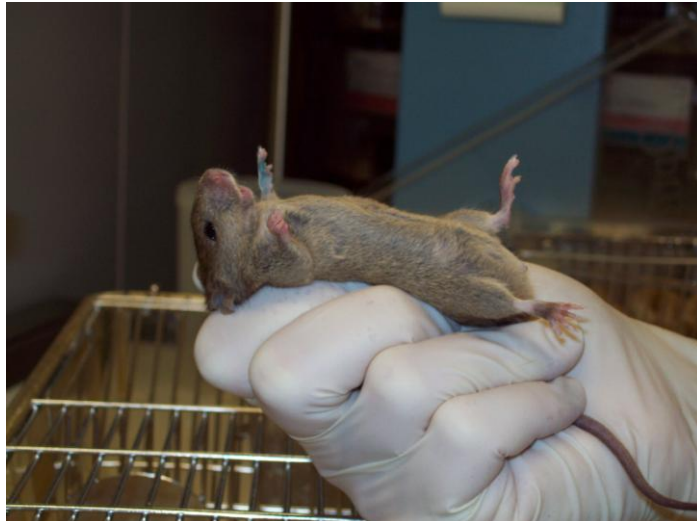
Ensure the mouse's head is facing and gently position the mouse so that its body is stretched out.

While holding the tail with one hand, use the other hand to grasp the loose skin from the nape of the neck to the base of the tail, with the thumb and forefinger.



Make sure to grasp enough scruff otherwise the mouse will be able to turn around and bite. Too much scruff and the airway can be restricted and the mouse can become cyanotic.

Monitor the animal closely the entire time it is restrained and gently release the animal if there are any signs of gasping or change in colour of the mucous membranes.



6. Mechanical restraint devices



When performing specialised techniques such as IV injections etc the mouse can be placed into a mechanical restraint device.

RECOMMENDATIONS:

- Method 1. Well handled animals prefer this method**
- Method 2. Do not leave the mouse to dangle – this can provoke the mouse to bite**
- Method 3. Recommended for heavily pregnant females**
- Method 4. Recommended for mice likely to bite when picked up by the tail**

Method 5. For procedures such as injections, blood collections and close examinations.

Method 6. For access to the mouse tail for intravenous injections and blood collections. Within UQ see a UQBR facility manager for further information and/or assistance.

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REVISED:

REFERENCES

1. <http://iws.ccccd.edu/biopage/mouse%20tutorial.pdf>
2. Hedrich, H., (ed.), *The Laboratory Mouse*, 656 pp. Elsevier Academic Press, New York, 2004. \$199.95. ISBN 0-12-336425-6.